# **Recent Technical Developments in the Standardized** Separation and Measurement of Extracellular Vesicles

#### Amy Phillips<sup>1</sup>, Robert Vogel<sup>2</sup>, and Murray F. Broom<sup>3</sup>

<sup>1</sup>Izon Science US Ltd, Christchurch, New Zealand (amy.phillips@izon.com), <sup>2</sup>The University of Queensland, Brisbane, QLD, Australia, <sup>3</sup>Izon Science Ltd, Christchurch, New Zealand

www.izon.com

#### The Need for Standardised Analysis In Extracellular Vesicle Research

The concentration, size, and charge of vesicles reflect the health of the organism or tissue they are extracted from<sup>1</sup>. Accurate measurement of these parameters therefore, has critical implications in the clinic. Furthermore, sample preparation can influence the vesicle characteristics<sup>2</sup>



A standardised method of vesicle preparation and analysis is needed to allow comparison between research groups.

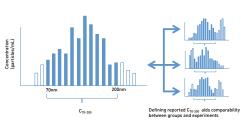
Current purification methods have problems with aggregation, or long, laborious processes. Commonly used characterization techniques lack sensitivity, and resolution3.

We have developed a method of isolation and characterization, which together provide a sensitive method for determining vesicle size and concentration.

#### Size-Based Concentration

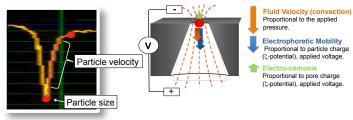
A new definition of particle concentration based on the size range being examined has been proposed, e.g. C70.200

This is more meaningful than the standard definition in that it specifies the range under consideration and of interest to the researchers (e.g. 70 200nm), and will aid comparability between measurements.



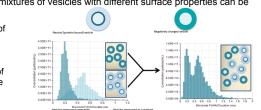
#### Particle Surface

Analysis of ζ-potential & surface modification



ζ-potential is calculated from the velocity of the particle as it moves through the nanopore. Changes to the particle surface through, for example, protein or aptamer binding can cause changes in  $\zeta$ -potential. Due to the particle-by-particle measurement, mixtures of vesicles with different surface properties can be measured

Surface charge of vesicles decreases vesicles protein binding. Ratios of differently charge vesicles can be detected.



### **TRPS: Detailed Size, Charge and** Concentration of EVs

Izon TRPS use a tunable elastic pore to measure particle properties. The size, charge and concentration of vesicles are determined with high resolution and sensitivity from the ionic current pulse signal they generate as they pass through the elastic pore4.



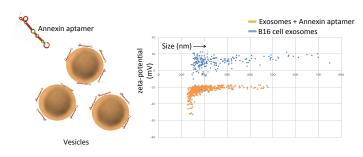


Tuning the pore size, applied pressure and voltage improves the measurement sensitivity and enables a wider range of particles to be analysed.

Particle Size: 50nm – 20µm+ Conc. Range: 105 - 1012 particles/mL Sample Size: 30uL Sample Media: Aqueous Buffer

## EV Identification

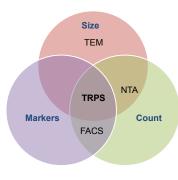
Aptamer based tool kit in development, to be used in conjunction with gNano to provide biological information.



The interaction of aptamers with biological targets on the vesicle surface, e.g. annexin, can provide discriminatory information based on the size and charge profiles of the different populations.

## Conclusions

- TRPS is the only high resolution technique which measures Size and Count, as well as surface Biomarkers (Figure)
- Size-Based Concentration is essential for meaningful comparison of measurements
- gEV columns provide a simplified approach to EV separation and isolation, and are expected to become the standard method



Comparison of the capability of techniques for analysing vesicles.

#### References

- van der Pol, et al., (2012) Classification, functions, and clinical relevance of extracellular vesicles, Pharmacol Rev 64, 1-33 Sowery et al., (2007) Dynamically resizable nanometer-scale apertures for molecular sensing, Sensors & Act B 123, 325-330 Yuana et al., (2011) Pre-analytical and analytical issues in the analysis of blood microparticles, Thromb Haemost 105, 396-408 Anderson et al., (2013) A comparative study of submicron particle sizing platforms: Accuracy, precision and resolution analysis of polydisperse particle size distributions, J Coll Inter Sci 405, 322-330 4. Roberts et al., (2012) Biosensors and Bioelectronics 31, 17-25